

wgMLST

Whole genome multi-locus sequence typing

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Introduction



Goals

Goals

Dataset

Credits

Disclaimer

- The wgMLST analysis strategy
- Why you should like the wgMLST approach
- An automated pipeline for wgMLST
- Analysis of *Listeria monocytogenes* surveillance data using wgMLST

Introduction



- Goal Dataset: real-time *Listeria* surveillance data
- Dataset
- 349 samples (356 sequencer runs) of *Listeria monocytogenes*
 - Clinical and environmental samples
- Credits
- Data set contains both sporadic samples and outbreak samples
- Disclaimer
- In particular, the data set contains data from the Crave Brothers Cheese outbreak.

Introduction



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- Data generation: CDC
- Sequencing platforms: Illumina (GAII, MiSeq)

Credits

- Data analysis: BioNumerics® 7.5 (Applied Maths)

Disclaimer

Introduction



Goal

Disclaimer

Dataset

For the last 7 years, I was affiliated with the following organization:

Credits

Disclaimer

- Organization:

Applied Maths NV, Keistraat 120, B-9830 Sint-Martens-Latem, Belgium

- Relationship:

employee (salary)

wgMLST



- Starting point : a set of loci
 - A locus is determined by
 - a set of variants for this locus, and
 - possibly, a set of validations for this locus (*in silico* PCR primers)
- For each sample, determine for each locus whether
 - the locus is present, and
 - if present, which variant (“allele”) of the locus is present
- Two procedures: Identify alleles
 - from (de novo) assembled genomes
 - directly from the reads

Allele identification



Two procedures:

- Assembly-based:

Identify alleles from (de novo) assembled genomes using BLAST

- ✓ Computationally intensive
- ✓ Multiple contigs lead to loci that are missing from the assembly
- ✓ Undefined behavior for multi-copy loci
- ✓ Required for extrinsic validation

- Assembly-free:

Identify alleles directly from the reads

- ✓ Computationally less intensive
- ✓ Missing loci are missing in the reads, not in the assembly
- ✓ Exhaustive procedure for multi-copy loci
- ✓ No extrinsic validation possible

Strong points



- ✓ Based on the concept of allelic variation, not only point mutations

Different estimate of evolutionary distance for events such as

- recombination
- simultaneous close-range mutations

are counted as one event, which might biologically more relevant

- ✓ Fixed set of loci leading to typing schemes on different levels



- ✓ Close to functional analysis
- ✓ Naturally incorporates multiple references
- ✓ Can be translated to SNPs within loci

Drawbacks



- ✓ Does not consider the “complete” genome
By default, intergenic regions only

- ✓ Requires curation

Since the goal is to have a universal set of loci and alleles that can be communicated about, consistent naming is required.

wgMLST pipeline



BioNumerics client



Isolate
database

External storage
NCBI, ENA, BaseSpace



BIGSdb
Public nomenclature



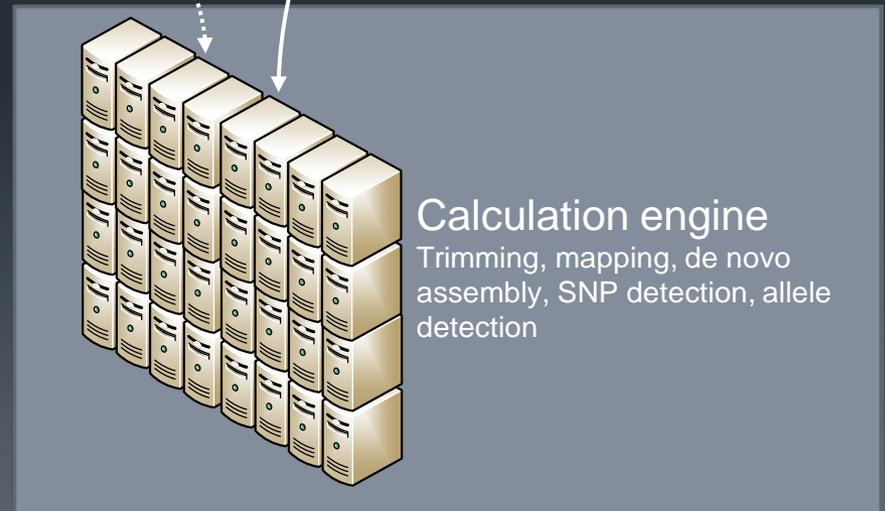
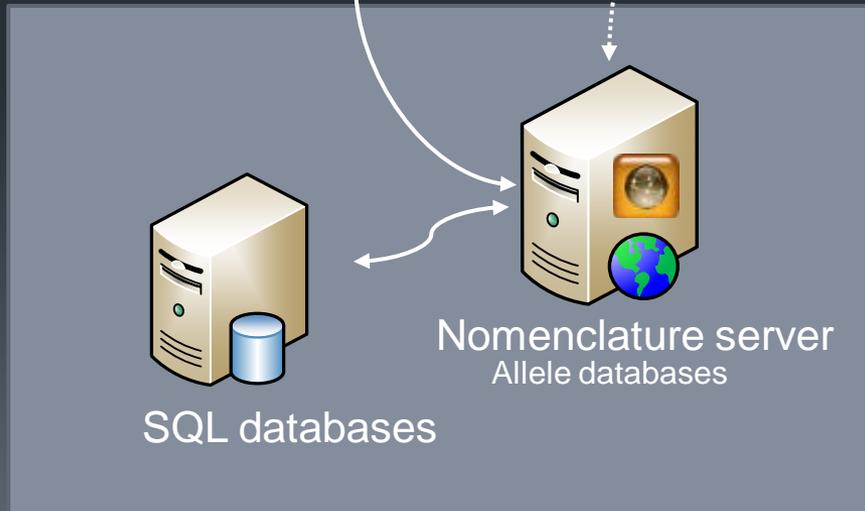
Nomenclature server
Allele databases



SQL databases



Calculation engine
Trimming, mapping, de novo
assembly, SNP detection, allele
detection



wgMLST pipeline



Sample side: lightweight sample database

- No heavy lifting in storage or in calculations
- Metadata remains local
- Total automation: “back to biology”
 - synchronized with NCBI BioProject
 - jobs are posted and retrieved automatically

wgMLST pipeline



Curation side: automated curation tools

- Alleles are automatically named (independent of public nomenclature)
validation criteria: deviation from closest known allele, check for start/stop codon, ...
- Automated sequence type assignment
- Automatic synchronization with public nomenclature
- Annotation of alleles, loci, sequence types, typing schemes ... used in sample reporting.

wgMLST pipeline demo



- Complete setup running at the Applied Maths booth
- Sample and allele databases available for
 - *Campylobacter*
 - *Listeria monocytogenes*
 - *Mycobacterium tuberculosis*
 - *Salmonella*
- New schemes need about a day to develop

Lmo wgMLST scheme



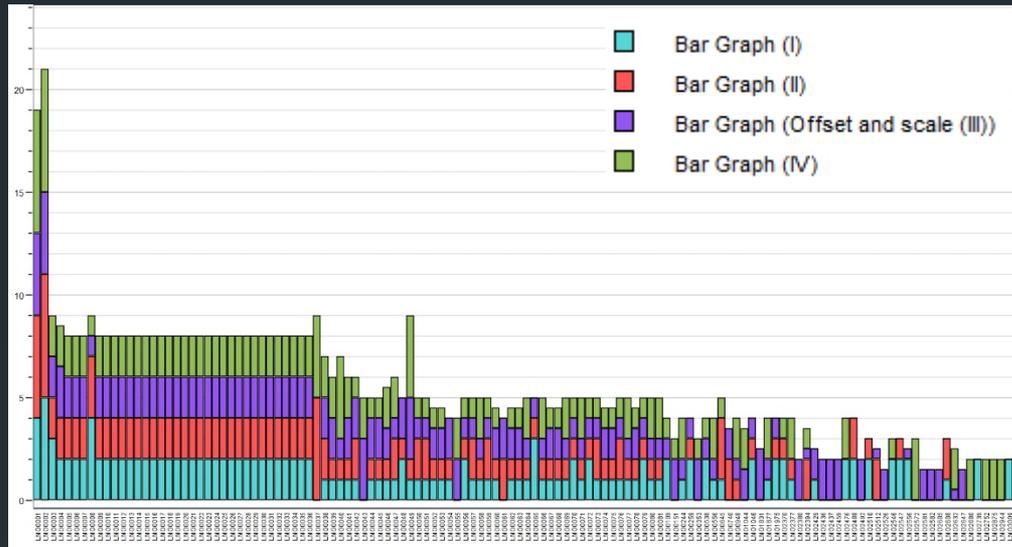
- 5 Reference sequences representing 4 lineages
FM242711, NC_003210, FM211688, CP001175, NZ_CM001469
- CDS's of these 5 sequences used to obtain homologous groups (Deng2010) and thus a set of well-defined loci.
 - Sequence identity > 50%
 - E-value < 0.00001

Lmo wgMLST scheme



- Number of loci: 3436
- Number of core loci: 2345 68.25%
(note: core = present in all lineages)
- Number of lineage-specific loci: 649 18.9%
(I: 134, II: 283, III: 156, IV: 76)
- Number of multi-allelic loci: 126 3.7%
(I: 56, II: 53, III: 74, IV: 66)

Lmo wgMLST scheme



- up to 6 copies of the same locus per sample
- some loci tend to be multi-copy in all lineages
- some loci are lineage-specific and multi-copy

PCR validations (20 bp length): largely lineage-specific

- Unique set of PCR primers: 779 loci (22.7%)
- Preceding PCR primer: 1147 loci (33.4%)
- Succeeding PCR primer: 976 loci (28.4%)

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