

Subtyping the top 30 *Salmonella* serotypes using a combination of CRISPR elements and virulence genes:

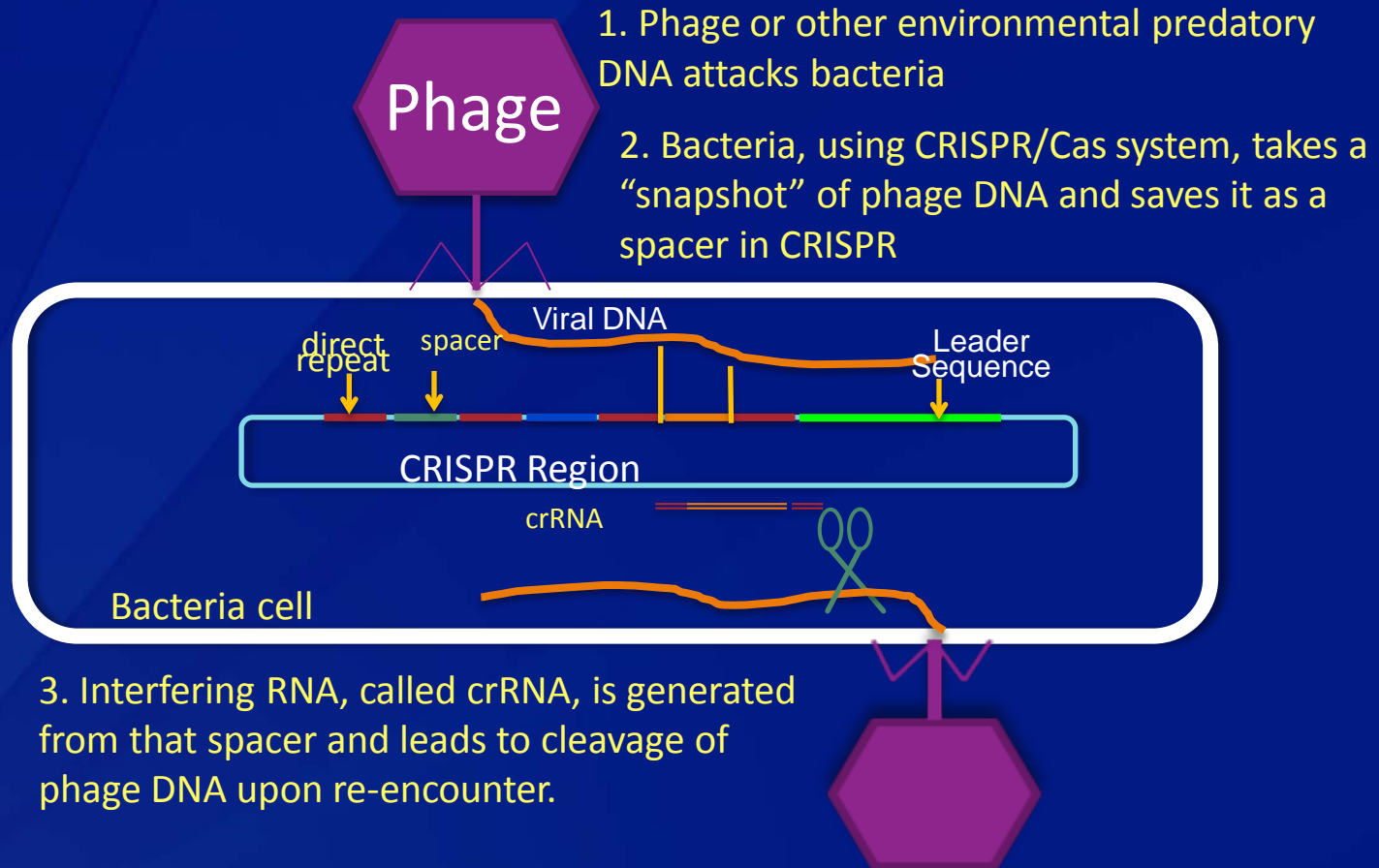
Salmonella CRISPR-MLVST

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Objectives

- ❑ Introduce the CRISPR/cas system
- ❑ Present subtyping scheme combining CRISPR and virulence genes (CRISPR-MLVST)
- ❑ Preliminary Data
- ❑ Challenges and Future Activities

Overview of CRISPR/Cas System



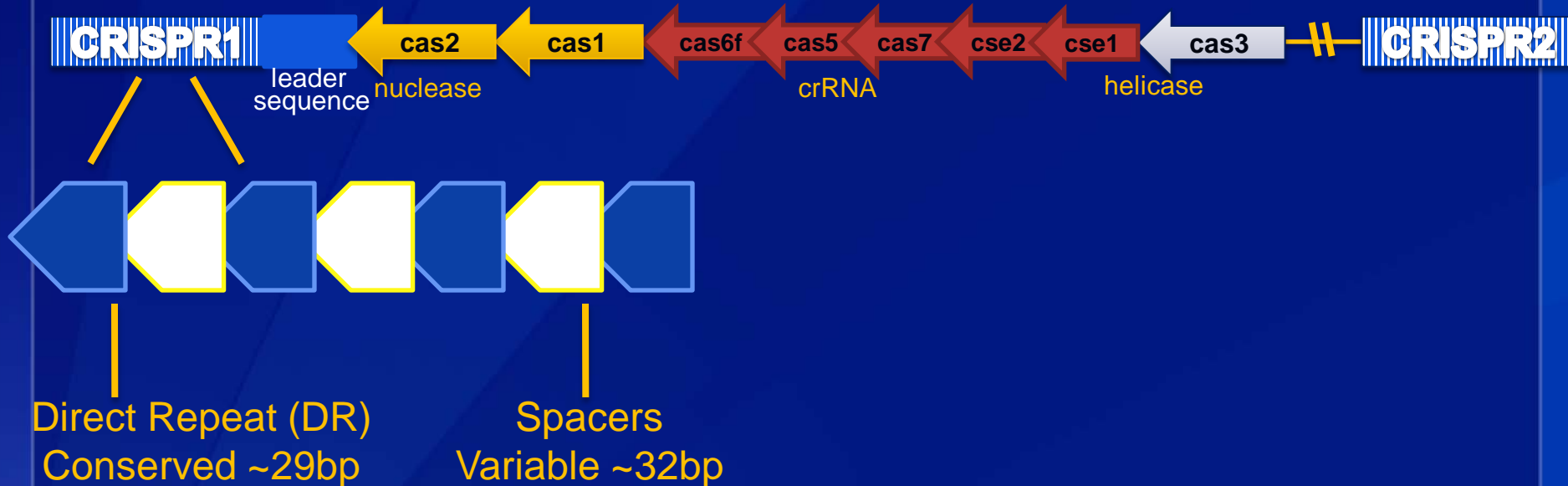
CRISPR/Cas System History

- ❑ First described in 1987 by Ishino et al in *E. coli*.
- ❑ “CRISPR” coined in 2000 by Mojica et al.

	Genomes Analyzed	CRISPR Loci Found
Archaea	150	557 (125)
Bacteria	2480	3355 (1126)
Total	2630	3912 (1251)

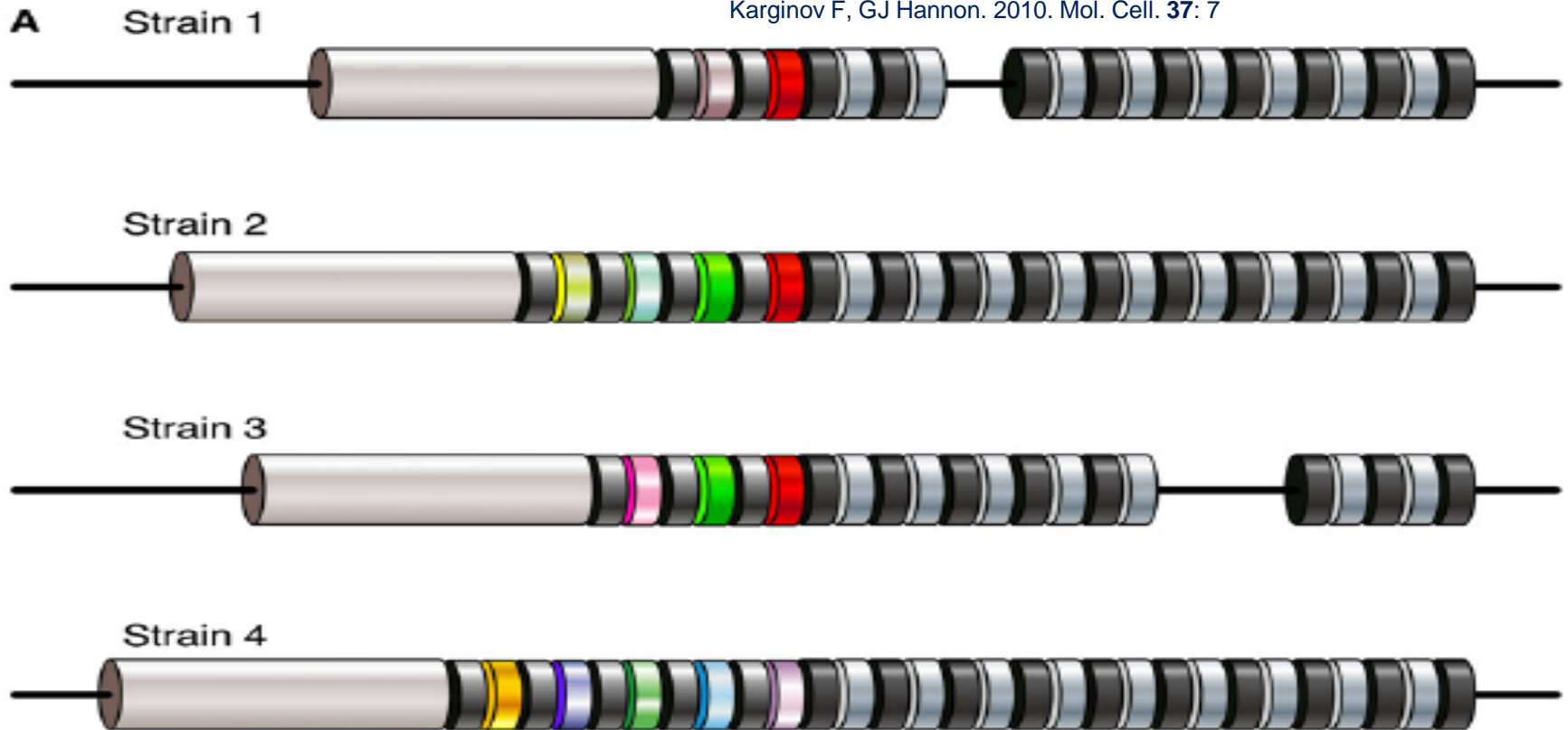
- ❑ CRISPR studied for subtyping different bacteria:
 - *Salmonella enterica*
 - *Campylobacter jejuni*
 - *Escherichia coli* and *Shigella*
 - *Vibrio cholerae*
- ❑ Level of discrimination depends on organism

CRISPR/Cas System



- ❑ New spacers added next to leader sequence
- ❑ Spacers a reflection of predatory DNA in environment and can be used for subtyping – bacteria from same environment will have same spacers

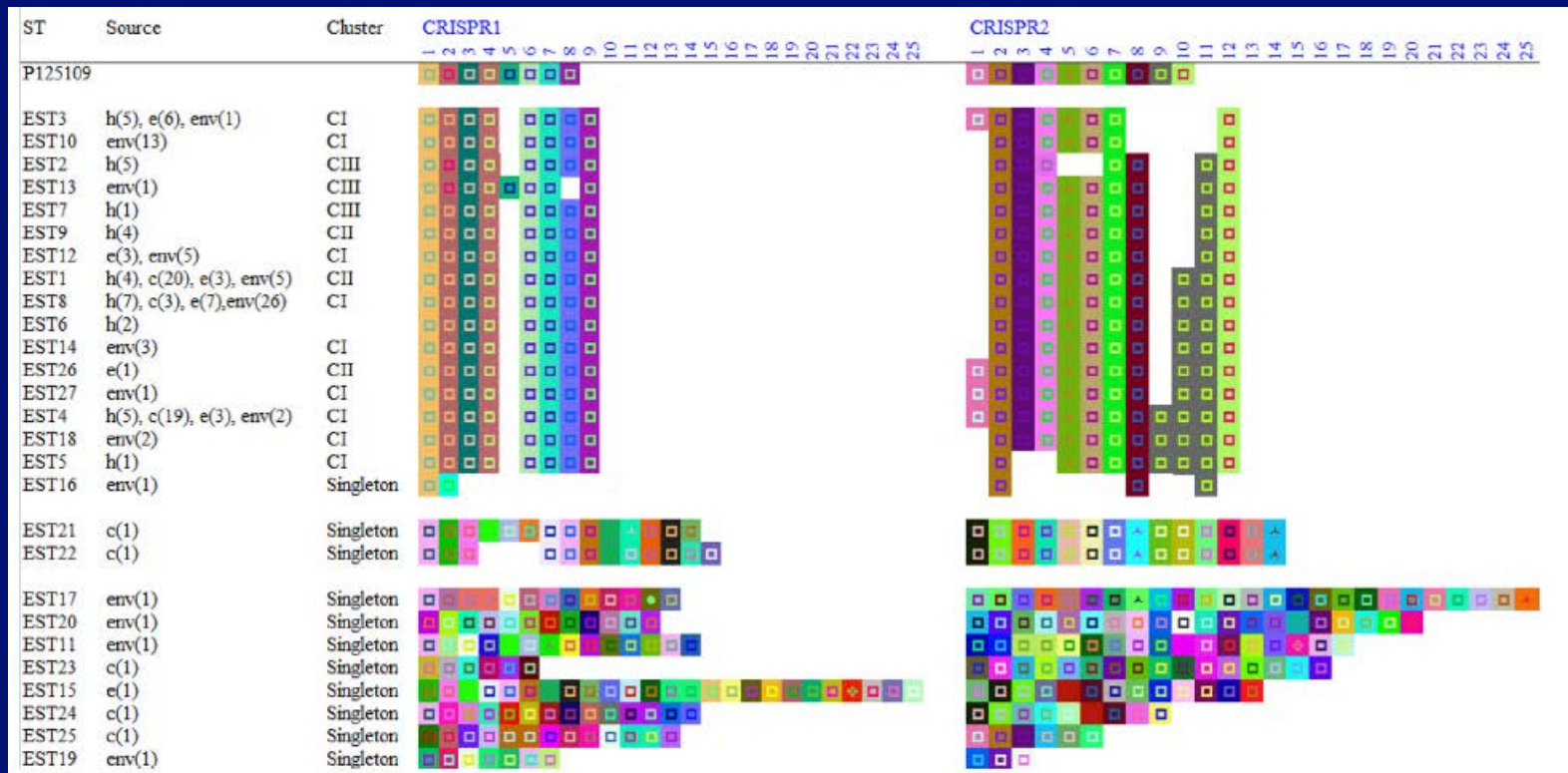
Subtyping using CRISPR region



- Subtyping based on the addition and deletion of spacer regions and variation between spacers

Subtyping using CRISPR Regions in *Salmonella*

- ❑ *Salmonella* has 2 CRISPR regions
- ❑ Ability to discriminate between serotypes is limited

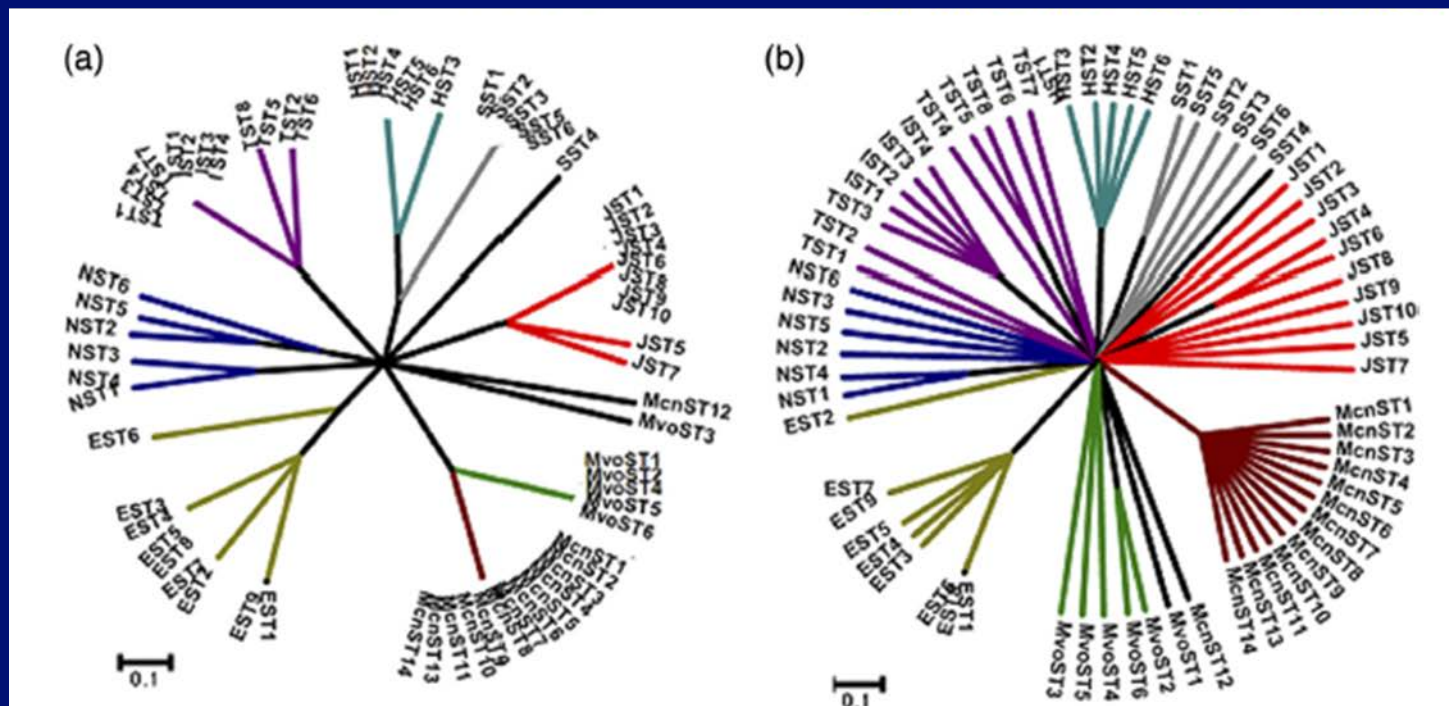


Subtyping with CRISPR and Virulence Genes in *Salmonella*

- Virulence genes group isolates by serotype while CRISPR 1 and 2 identify clusters of epidemiologically related isolates within a serotype

fimH and *sseL*

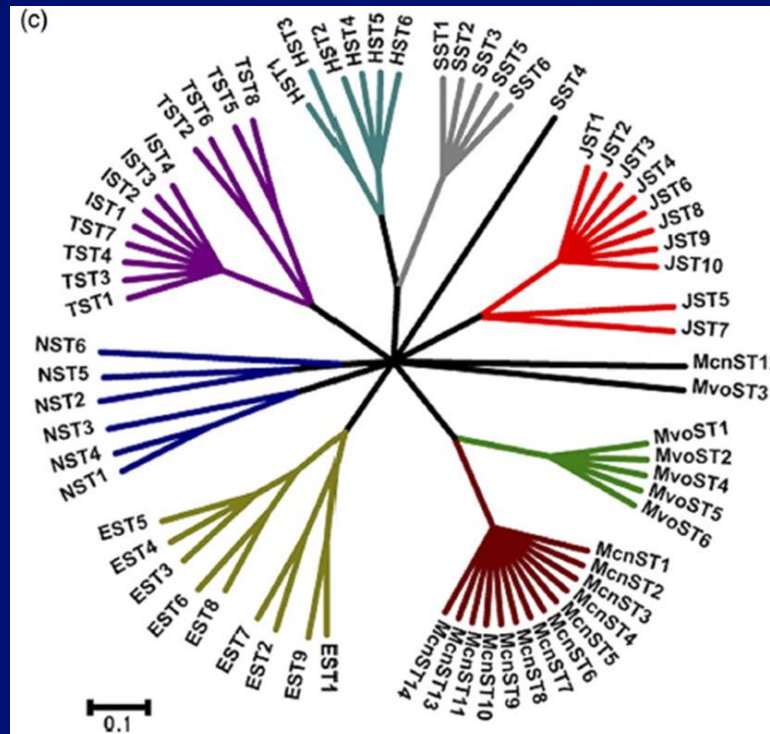
CRISPR1 and CRISPR2



Salmonella CRISPR-MLVST

- Virulence genes group isolates by serotype while CRISPR 1 and 2 identify clusters of epidemiologically related isolates within a serotype

sseL, *fimH*, and CRISPR1-CRISPR2



***Salmonella* CRISPR-MLVST study design**

- ❑ Perform CRISPR-MLVST on both sporadic and epidemiologically associated isolates from top 30 serotypes of *Salmonella enterica*
- ❑ Determine whether this method is comparable to other subtyping methods including PFGE and MLVA

Methods for CRISPR-MLVST

- ❑ Perform Sanger sequencing on all 4 targets using previously designed primers from Penn State collaborators and Pasteur Institute Primers
- ❑ Develop a database using the Bionumerics software package to:
 - Assign CRISPR types based on variable spacer regions only
 - Assign allele types to the virulence genes *sseL* and *fimH*

Assigning Allele Numbers

- ❑ **For MLVST: A new allele is defined as at least single base pair difference in the coding region for *sseL* or *fimH* that has not been previously identified**
- ❑ **For CRISPR1 and 2:**
 - New spacer is defined as at single base pair difference from previous identified spacers
 - New allele is a new spacer order not previously identified
- ❑ **Overall sequence type based on all four targets and is a combination of all 4 allele numbers (*fimH:sseL:CRISPR1:CRISPR2*)**

Preliminary Results

Category	Number
Total number of isolates	958
Total number of serotypes	32
Number of <i>fimH</i> alleles	68
Number of <i>sseL</i> alleles	76
Number of CRISPR 1 alleles	189
Number of CRISPR 2 alleles	134
Number of isolates sequenced at all 4 alleles	106

CRISPR 1 Region

- ❑ 43 different repeat sequences identified ranging from 28-29 bp
- ❑ 478 different spacers identified ranging from 11-64 bp (average 32bp)
- ❑ Number of spacers per allele: range 2 to 32



CRISPR 2 Region

- ❑ 48 direct repeats identified ranging from 26-30bp (average 29)
- ❑ 547 spacers identified that were 28-54bp long (average 32bp)
- ❑ Number of spacers per allele: range 2 to 26

α2_032 α2_064 α2_001 α2_061 α2_001 α2_032 α2_001 α2_076 α2_001 α2_070 α2_001 α2_071 α2_001 α2_002 α2_001 α2_078 α2_001 α2_074 α2_001 α2_065 α2_001 α2_035 α2_001 α2_076 α2_001 α2_094 α2_001 α2_031 α2_001

***fimH* and *sseL* Alleles**

- ❑ **Most common differences between isolates are single nucleotide polymorphisms**
- ❑ **Most serotypes only have 1-2 allele types**

CRISPR-MLVST Results for Serotype Saint Paul: Low Epidemiological Concordance

<i>fimH</i>	<i>sseL</i>	CRISPR1	CRISPR2	Oubreak associated
17	20	28	39	0806MAJN6-1c
17	54	105	45	Sporadic
17	20	28	130	Sporadic
17	20	20	134	Sporadic
17	20	20	31	Sporadic
17	20	20	31	0805NMJN6-1c
17	20	20	31	0805NMJN6-1c
17	20	27	40	Sporadic
17	20	27	40	0902NEJN6-1
17	20	27	40	0902NEJN6-1
17	20	27	40	0902NEJN6-1
17	20	20	41	Sporadic
17	20	20	41	Sporadic
17	20	20	41	Sporadic
17	20	20	41	Sporadic
17	20	20	41	0805NMJN6-1c
17	20	20	41	0805NMJN6-1c
17	20	20	41	0805NMJN6-1c
17	20	28	42	0805NMJN6-1c

CRISPR-MLVST Results for Serotype Newport: High Epidemiological Concordance

<i>fimH</i>	<i>sseL</i>	CRISPR1	CRISPR2	Outbreak-associated
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	6	126	11	1005SDCJJP-1
5	8	7	12	1008MDJJP-1
5	8	7	12	1008VAJJP-1
5	8	7	12	1008VAJJP-1
5	6	103	11	1005UTJJP-1
5	6	103	11	1005UTJJP-1
3	5	90	10	Historical isolate

Challenges

- ❑ **Difficult to design universal primers for all *Salmonella* serotypes**
- ❑ **The same primer design does not work for all isolates within the same serotype**
- ❑ **CRISPR region variation and size**
 - As more spacers are added the CRISPR region becomes larger, pushing the limits of what we can feasibly Sanger sequence
 - CRISPR regions vary requiring design of serotype specific primers

Future Plans

- Characterize additional isolates at all four targets to further understand diversity at CRISPR and virulence genes**
- Include CRISPR regions and virulence genes as part of the genetic information extracted from whole genome sequences as use of NGS increases**

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